



## Both ozone exposure and soil water stress are able to induce stomatal sluggishness

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### ABSTRACT

We tested whether short-term exposure to realistic ozone pollution ( $\leq 150$  ppb, 1 h) and soil water stress (soil water content  $\leq 15\%$ ) slow stomatal dynamics in an ozone-sensitive cultivar of snapbean. Both ozone exposure and water stress caused stomata to be sluggish in the degree of closure after leaf severing, while ozone also delayed the time the closing signal was perceived. Ozone-induced aberrations lasted up to the night and caused incomplete closure of stomata. No synergic effect was observed in the dynamic measurements. In contrast, at steady-state, water stress protected the plants from the negative ozone effects on stomatal conductance. Ambient ozone peaks may thus cause sluggish stomatal response and increase leaf water loss both under well watered and drought conditions.

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### 1. Introduction

Tropospheric ozone ( $O_3$ ) is an important secondary pollutant resulting from emission of volatile organic compounds and nitrogen oxides, and is also recognized as a significant greenhouse gas (Bytnerowicz et al., 2007; Serengil et al., 2011). Ozone is seriously phytotoxic and causes negative effect on plants (e.g., NIES, 1980, 1984; Omasa et al., 2002; Paoletti, 2007). Stomatal  $O_3$  uptake is crucial for assessing the adverse effect of  $O_3$  on plants (Omasa et al., 1979; UNECE, 2004; Paoletti and Manning, 2007; Grulke et al., 2007a; Cieslik et al., 2009). However, our understanding about stomatal responses to  $O_3$  is still imperfect (Paoletti and Grulke, 2005). Previous studies reported that  $O_3$  generally induces both stomatal closure (Wittig et al., 2007, 2009) and sluggishness of stomatal response to change of environmental factors (Paoletti and Grulke, 2005, 2010). Because plants live in a fluctuating environment, both steady-state stomatal conductance and stomatal dynamics play an important role in regulating leaf gas exchange. Ozone-induced sluggishness of stomata has been reported in response to change in photosynthetic photon flux density (PPFD) (Reich and Lassoie, 1984; Reiling and Davison, 1995; Paoletti, 2005; Grulke et al., 2007a; Paoletti and Grulke, 2010), vapor pressure deficit (VPD) (Grulke et al., 2007b) and severe water stress imposed by severing a leaf (Paoletti, 2005; Paoletti et al., 2009; Mills et al., 2009). Such aberrations may increase nighttime transpiration, as reported for

several tree species exposed to daytime  $O_3$  exposure in controlled (Skärby et al., 1987; Matyssek et al., 1995; Wieser and Havranek, 1995; Günthardt-Goerg et al., 1997; Grulke et al., 2007c) and ambient conditions (Grulke et al., 2004). Ozone is known to induce up-regulation of ethylene emission, which has been suggested to be responsible for a reduction in stomatal sensitivity to abscisic acid (ABA) and thus to closure (Wilkinson and Davies, 2010). Several stressors, including water deficit, are known to stimulate ethylene production by plants (Morgan and Drew, 1997).

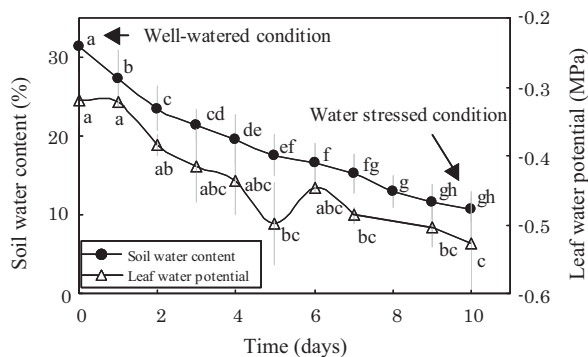
Climate change is expected to increase  $O_3$  levels and alter precipitation regimes, regionally increasing the probability of drought (Ciais et al., 2005). Drought has the capacity to limit  $O_3$  injury through stomatal closure (Tingey and Hogsett, 1985), while  $O_3$ -induced impairment of stomatal response may increase susceptibility to drought (Nali et al., 2004).

Our main objective was to test whether short-term exposure to realistic  $O_3$  pollution and soil water stress, singly and in combination, may slow stomatal dynamics in an ozone-sensitive cultivar of snapbean (*Phaseolus vulgaris*, S156).

### 2. Materials and methods

Seeds of the ozone sensitive cultivar S156 of snapbean developed at the Raleigh USDA-ARS (Burkey and Eason, 2002; Flowers et al., 2007; Booker et al., 2009), were planted in 17-cm (1.7–1) pots, filled with sand:peat:soil = 1:1:1 (v:v:v). Seed were planted over several days so that same-age (4-week old) plants were used in the experiment. All plants were grown in a room with controlled environmental conditions (air temperature of 20 °C, PPFD

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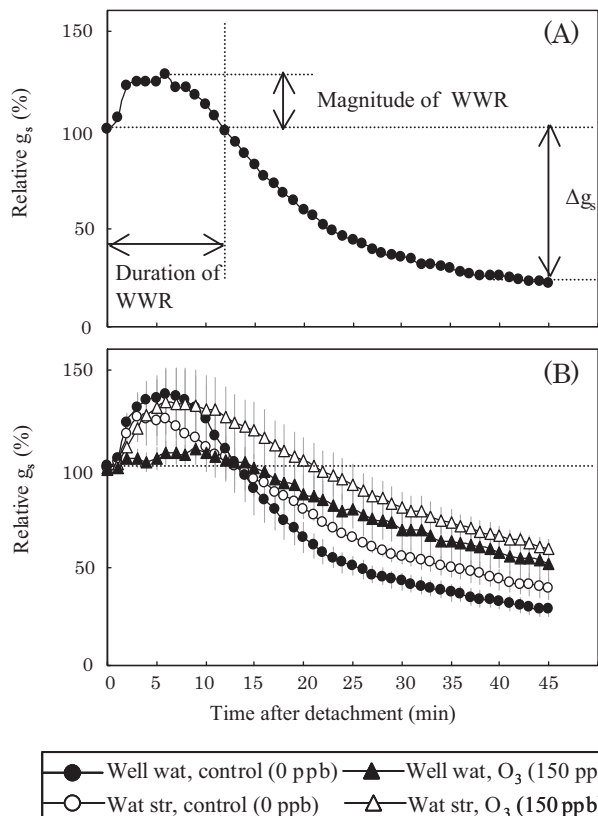
**Fig. 1.** Changes in soil water content ( $\pm$ S.D.,  $n=18$ ) (%) and predawn leaf water potential ( $\pm$ S.D.,  $n=2-3$ ) after irrigation until field capacity. Different letters indicate significant differences among mean values (Tukey HSD test,  $P < 0.05$ ).

of  $500 \mu\text{mol m}^{-2} \text{s}^{-1}$ , and relative humidity of 55%). Well-watered plants were supplied with water every day and in particular immediately before the ozone exposure, while water-stressed plants received no irrigation for 10 days before  $\text{O}_3$  fumigation. Soil water content was more than 30% in well-watered plants and 6–15% in water-stressed plants, corresponding to a predawn leaf water potential of  $-0.32$  MPa and  $-0.53$  MPa, respectively (Fig. 1). Water content was measured in 18 pots with an ECH2O EC-5 soil moisture sensor (Decagon Devices, Pullman WA, USA) at root level. Leaf water potential was measured at dawn in a selection of 2–3 plants per day by means of an SKPM 1400 chamber pressure (Skye, Powys, UK).

The fully expanded central leaf of the second trifoliate leaf was selected as a target leaf. After 1-h exposure to  $1000 \mu\text{mol m}^{-2} \text{s}^{-1}$  light, steady state leaf gas exchange was measured with a portable infra-red gas-analyzer (CIRAS-2 PP Systems, Herts, UK), equipped with a  $2.5 \text{ cm}^2$  leaf cuvette which controlled leaf temperature ( $20^\circ\text{C}$ ), leaf-to-air vapour pressure deficit ( $0.9 \text{ kPa}$ ), saturating light ( $1800 \mu\text{mol m}^{-2} \text{s}^{-1}$ ) and  $\text{CO}_2$  concentration ( $365 \text{ ppm}$ ). Ozone exposure to one of four levels ( $\pm$ SD), i.e. low ( $48 \pm 6.7 \text{ ppb}$ ), middle ( $87 \pm 9.4 \text{ ppb}$ ), high ( $150 \pm 10.9 \text{ ppb}$ ), and control ( $0 \text{ ppb}$ , no  $\text{O}_3$ ), was then carried out for 60 further min by a web fumigation system (Velikova et al., 2005; Pinelli and Tricoli, 2008). Ozone was added by an  $\text{O}_3$  generator (Model Heliozon, Milano, Italy) to the fumigating air for the target leaf through a teflon tube. The concentration around the leaf was recorded with an ozone monitor (Mod. 205, 2B Technologies, Boulder CO, USA), and adjusted through mass flow controllers (Mod. GFC171S Aalborg). At 30 min after the end of  $\text{O}_3$  exposure, steady-state stomatal conductance ( $g_s$ ) was measured again. When  $g_s$  reached equilibrium under constant light at  $1800 \mu\text{mol m}^{-2} \text{s}^{-1}$ , the methodology described by Paoletti (2005) was applied to assess dynamic variations of  $g_s$  after cutting the leaf petiole (Fig. 2). Data were logged at 1 min intervals in the 45 min after severing. In the experiment, two phases of  $g_s$  response were observed (Fig. 2). At first,  $g_s$  increased for Iwanoff effect. This transient increase called as the transient 'wrong-way response' (WWR) is due to a difference in turgor pressure between guard cell and epidermal cells (Omasa and Maruyama, 1990; Powles et al., 2006). Subsequently, stomatal conductance decreased with increasing leaf water stress. WWR duration and magnitude of WWR and  $g_s$  decrease at 45 min ( $\Delta g_s$ ) were recorded.

After  $\text{O}_3$  exposure and steady-state measurements, 22 plants were placed in the dark for 10 h. Nocturnal steady-state  $g_s$  was measured with leaf temperature of  $20^\circ\text{C}$ , leaf-to-air vapour pressure deficit of  $0.9 \text{ kPa}$ , no exposure to light and  $\text{CO}_2$  concentration of  $365 \text{ ppm}$ .

Data were checked for normal distribution and homogeneity of variance (Levene's test). Percents were arcsine square root



**Fig. 2.** Time courses of stomatal conductance ( $g_s$ ) after severing a leaf at time 0. Graph A shows an example of calculation of WWR magnitude and duration, and of the degree of  $g_s$  decrease ( $\Delta g_s$ ). At first,  $g_s$  showed a transient increase called 'wrong-way response' (WWR) and a subsequent decrease with increasing leaf water stress. Graph B shows the time course of average  $g_s$  ( $\pm$ SE) under well-watered condition (soil water content  $> 30\%$ ) and water-stressed condition (soil water content = 6–15%) in control ( $0 \text{ ppb O}_3$ ) and ozone-exposed ( $150 \text{ ppb O}_3$ ) leaves.

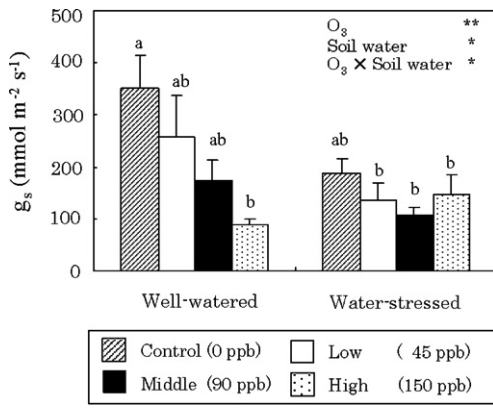
transformed prior to analysis. Effects of soil water status and  $\text{O}_3$  exposure were tested using two-way analysis of variance (ANOVA). Results were considered significant at  $p < 0.05$ . Differences among means were tested by Tukey's HSD test. Statistical analysis was performed with STATISTICA software (6.0, StatSoft Inc., Tulsa, OK, USA), according to Statsoft (2001).

### 3. Results

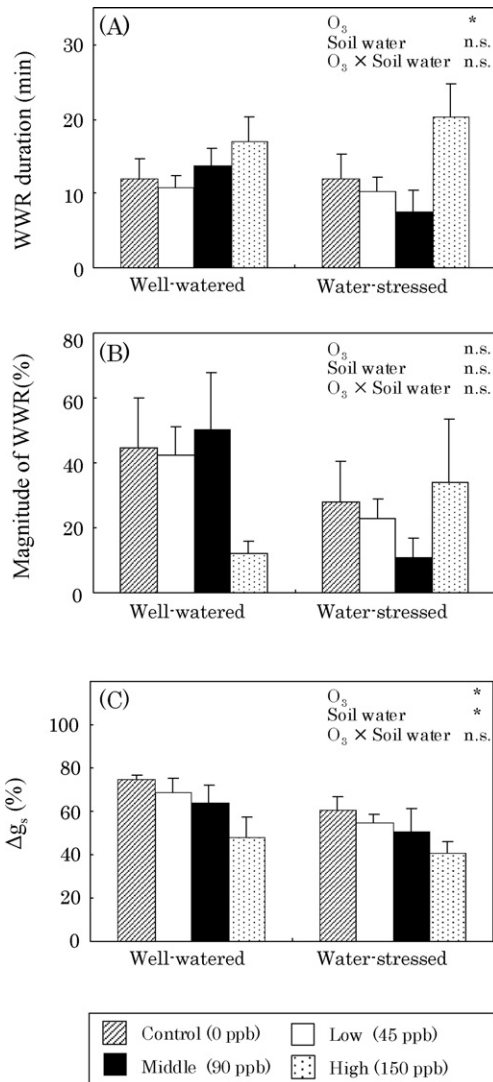
#### 3.1. Steady-state and dynamic stomatal responses

Ozone exposure induced a decline of steady-state  $g_s$  under well-watered conditions (Fig. 3).  $g_s$  was 73% in the low  $\text{O}_3$  treatment and 25% in the high  $\text{O}_3$  treatment compared to the control plants. A reduced soil water availability significantly reduced  $g_s$  relative to well-watered plants and resulted in no effect of  $\text{O}_3$  on  $g_s$ .

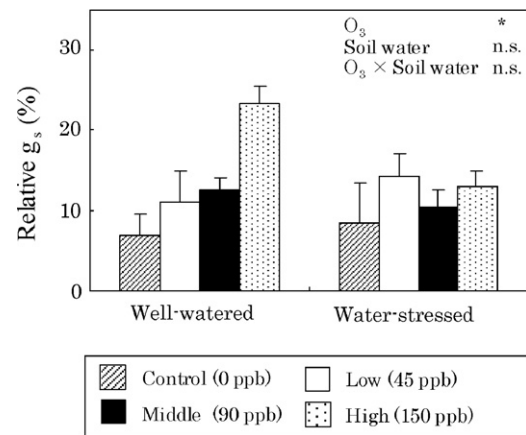
After severing a leaf, two phases of  $g_s$  response were observed (Fig. 2): a transient increase as WWR and then a linear decrease. Ozone exposure increased WWR duration from  $11.9 \pm 2.7 \text{ min}$  in the control plants to  $17.1 \pm 3.2 \text{ min}$  in the high  $\text{O}_3$  treatment, while the effect of water stress was not significant (Fig. 4A). Magnitude of WWR was not affected by both  $\text{O}_3$  and soil water availability (Fig. 4B). Ozone reduced the degree of stomatal closure over time ( $\Delta g_s$ ) in both soil water conditions (Fig. 4C). Also soil water deficit reduced  $\Delta g_s$  relative to the optimal soil water availability. In well-watered plants, the high  $\text{O}_3$  treatment resulted in smaller stomatal closure than in control leaves (48% vs. 75%). In



**Fig. 3.** Effect of 1-h exposure to four levels of O<sub>3</sub> (control: 0 ppb; low: 45 ppb; middle: 90 ppb; high: 150 ppb) and soil water availability (well-watered 30% water content; water-stressed: 6–15% water content) on steady-state stomatal conductance ( $g_s$ ). Data are means ( $N=5-8$ )  $\pm$  SE. \* denotes significance at 5% levels; \*\* denotes significance at the 1% levels; n.s. indicates no significance. Different letters indicate significant differences among bars (Tukey HSD test,  $P < 0.05$ ).



**Fig. 4.** Effects of 1-h O<sub>3</sub> exposure (Control: 0 ppb; low: 45 ppb; middle: 90 ppb; high: 150 ppb) and soil water availability (well-watered 30% water content; water-stressed: 6–15% water content) on dynamic  $g_s$  response after severing a leaf (A: duration of transient increase in  $g_s$  after severing; B: magnitude of transient increase in  $g_s$  after severing; C: degree of  $g_s$  decrease at 45 min after severing). Data are means ( $N=3-4$ )  $\pm$  SE. \* denotes significance at the 5% levels; n.s. indicates no significance.



**Fig. 5.** Effects of 1-h O<sub>3</sub> exposure (Control: 0 ppb; low: 45 ppb; middle: 90 ppb; high: 150 ppb) and soil water availability (well-watered 30% water content; water-stressed: 6–15% water content) on nocturnal steady-state stomatal conductance ( $g_s$ ) relative to the pre-exposure daytime value. Data are means ( $N=2-4$ )  $\pm$  SE. \* denotes significance at the 5% levels; n.s. indicates no significance.

water-stressed plants, high O<sub>3</sub> and control treatment resulted in 41% and 61% stomatal closure, respectively.

### 3.2. Nocturnal stomatal conductance

Fig. 5 shows nocturnal steady-state  $g_s$  relative to daytime pre-exposure  $g_s$ . Ozone exposure caused an increase of nocturnal  $g_s$ . A reduced soil water availability did not affect nocturnal  $g_s$ . Nocturnal  $g_s$  increased in well-watered plants from 7% in the control plants to 23% in the high O<sub>3</sub> treatment.

## 4. Discussion

Increasing O<sub>3</sub> exposure under optimal water availability progressively decreased steady-state  $g_s$ . Although the mechanisms that regulate stomatal responses to acute and chronic O<sub>3</sub> exposure may differ, the unifying result is a reduction of steady-state  $g_s$ , in both crops and trees, under different experimental conditions (e.g., Grulke et al., 2007a; Wittig et al., 2007). Dynamic measurements of  $g_s$  in Arabidopsis, in contrast, showed a rapid decrease triggered by acute exposure (Vahisalu et al., 2010) and followed by reopening to overshooting values (Moldau et al., 2011). Soil water stress is usually considered to reduce O<sub>3</sub> injury because it reduces  $g_s$  and thus O<sub>3</sub> entering into a leaf (Tingey and Hogsett, 1985). The result of our steady-state measurements supports this conclusion. Simulated water deficit resulted in no effect of O<sub>3</sub> on  $g_s$  (Fig. 3). During the dry summer of 2003 in Central Europe, no difference in steady-state  $g_s$  was reported for beech trees exposed to ambient and twice-ambient O<sub>3</sub> in free air (Löw et al., 2006). Figures obtained from steady-state observations have been used to model plant responses to O<sub>3</sub> in a changing climate (e.g., Ollinger et al., 2002).

Measurements of dynamic stomatal responses revealed that exposure to increasing O<sub>3</sub> concentrations made stomata sluggish i.e. progressively increased the duration of WWR and reduced the degree of stomatal closure over time (Fig. 4A and C). After leaf severing, duration of WWR and following linear reduction in  $g_s$  are related to induction and execution of guard cell osmoregulation, respectively (Powles et al., 2006). Omasa (1990) reported that stomatal response was affected by the O<sub>3</sub>-induced effects such as slight increase in permeability of epidermal cell membranes and alteration of the osmotic pressure modulating a balance in turgor between the guard and subsidiary cells. Ozone may also delay stomatal responses by stimulating ethylene production and reducing



stomatal sensitivity to ABA (Wilkinson and Davies, 2010), or temporarily desensitizing the guard cells by blocking the  $K^+$  channels (Vahisalu et al., 2010).

We also demonstrated, for the first time that also a mild soil water stress is able to cause stomatal sluggishness by reducing the degree of stomatal closure, while no effect was recorded on the time the closing signal is perceived. Following Powles et al. (2006), this response suggests that water stress affects execution rather than induction of guard cell osmoregulation. In contrast with steady-state  $g_s$ , water deficit did not provide any protection from  $O_3$  effects on stomatal dynamics.

The effect of water stress on stomatal sluggishness, however, lasted shorter than the effect of ozone exposure. The imperfect stomatal closure at night, in fact, with higher nocturnal steady-state  $g_s$  after short-term  $O_3$  exposure in well-watered plants, may be considered as a long-lasting effect of the aberrations induced by ozone. Enhanced nocturnal  $g_s$  induced by  $O_3$  has been reported for several plant species (Skärby et al., 1987; Matyssek et al., 1995; Wieser and Havranek, 1995; Grulke et al., 2004, 2007c). Ozone-induced incomplete closure of stomata at night may translate into losing control of water efflux at dark (Skärby et al., 1987; Grulke et al., 2007c). Increase in nocturnal  $g_s$  may also enhance  $O_3$  uptake at night. Günthardt-Goerg et al. (1997) reported that considerable  $O_3$  uptake at night induced leaf injury in *Betula pendula*, *Populus × euramericana*, and *Alnus glutinosa*. Matyssek et al. (1995) reported that birch species exposed to  $O_3$  at night showed great reductions in growth.

Ozone peaks may exceed 0.1 ppm h in suburban and rural areas of California in the United States (Heath et al., 2009), southern Europe (Paoletti, 2006) and East Asia such as Japan (Takigawa et al., 2007). The present study revealed that realistic short-term  $O_3$  exposure ( $\leq 150$  ppb) induced stomatal sluggishness with or without drought stress. Although the effect is lighter than for ozone, also soil water deficit can induce stomatal sluggishness. Climate change brings about the risk of drought and flooding (Bytnerowicz et al., 2007). Ozone- and drought-induced loss of stomatal function may enhance both leaf water loss and  $O_3$  uptake. Current modeling efforts of  $O_3$  effects on plants have been developed using steady-state parameters (Emberson et al., 2000; Grünhage et al., 2001) and the  $O_3$ -induced losing control of dynamic stomatal response was ignored. The results presented here suggest to reconsider the role of  $O_3$  pollution on leaf gas exchange and highlight complex interactions between ozone and drought. Further improvement about our understanding of stomatal response to  $O_3$  and drought will contribute to assess climate change impacts on plant water balance and susceptibility to stress.

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